

**A MEMORY ENHANCING PROTEIN**

**CROSS REFERENCE TO RELATED APPLICATION**

This application claims priority from U.S.  
5 Provisional Application No. 60/198,802, filed April 20,  
2000.

**FIELD OF THE INVENTION**

The present invention provides methods for  
enhancing memory in animals, including humans. The  
10 present invention is also directed to methods for  
treating brain or spinal injury with the administration  
of an effective amount of atypical forms of protein  
kinase C, including protein kinase M zeta (PKM $\zeta$ ). The  
present invention further provides a method of inducing  
15 amnesia with the administration of an effective amount of  
a PKM $\zeta$  inhibitor.

**BACKGROUND OF THE INVENTION**

A common working hypothesis for the  
20 physiological basis of memory is that persistent changes  
in behavior are mediated by long-term modifications in  
the strength of synapses (Kandel et al. (1982) Science  
218:433-443; Bliss et al. (1993) Nature 361: 31-39). The  
molecular mechanisms for these changes are complex,  
25 involving many signal transduction pathways. Overall,  
however, these mechanisms are divided into two  
functionally distinct phases: induction, which initiates  
the long-term modifications, and maintenance, which  
sustains the changes (Malinow et al (1988) Nature  
30 335:820-824; Schwartz, J.H. (1993) PNAS 90:8310-8313;  
Schwartz et al (1987) Ann. Rev. Neurosci. 10:459-476).

Much of the work to examine these signaling

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pathways has come from the study of the response to high-frequency afferent stimulation of synapses that causes a long-term increase in synaptic transmission, long-term potentiation (LTP) (Bliss et al. (1993), *supra.*; Bliss et al. (1973) J. Physiol. 232:331-356; Nicoll et al. (1988) 1:97-103). The vast majority of signaling molecules implicated in LTP affect only induction, but not maintenance. The exceptions are agents that inhibit the catalytic domain of protein kinases, specifically protein kinase C (PKC), which are able both to block LTP induction and reverse its maintenance. (Nishizuka, Y (1988) Nature 334:661-665; Schwartz, J.H. (1993) *supra.*; Schwartz et al (1987) *supra.*

These two phases can be distinguished experimentally by the timing of the application of pharmacological agents that inhibit signal transduction pathways. When agents are applied prior to a tetanic afferent stimulation and prevent the formation of long-lasting changes, they block induction. If they are applied after the tetanus-and reverse the potentiation that has been established - they affect maintenance.

Several principles have been proposed to characterize mechanisms that might maintain long-term changes in synaptic transmission. First, protein kinases, such as PKC, which transiently enhance synaptic transmission when second messengers are activated, can extend their action by becoming constitutively active kinases that are independent of second messengers. (Schwartz et al (1987) *supra.*; Klann et al. (1991) J. Biol. Chem. 266:24253-24256)

Second, long-term forms of synaptic plasticity are thought to depend upon new protein synthesis, although the critical, newly synthesized molecules that cause synaptic potentiation have not been identified.

- 5 Stanton et al. (1984) J. Neurosci. 4:3080-3088; Frey et al (1988) Brain Res. 452:57-65; Otani et al (1989) Neurosci. 28: 519-526; Abel et al. (1998) Science 279: 338-341. A similar requirement for new protein synthesis has been observed for long-term memory. Davis et al.  
10 (1984) Psychol Bull. 96:518-559; Thompson, R.F. Science 233:941-947; Montarola et al. (1986) Science 234:1249-1254.

- While usually considered properties of separate mechanisms, it has been determined that one isoform of  
15 PKC possesses both of these features: it is persistently increased during LTP as a constitutively active enzyme, and it is generated by new protein synthesis. Sacktor et al. (1993) Proc. Natl. Acad. Sci. (USA) 90:8342-8346. This newly described form of PKC is PKM $\zeta$ , the independent  
20 catalytic domain of the PKC $\zeta$  isoform, which, lacking PKC $\zeta$ 's autoinhibitory regulatory domain, is autonomously active. Schwartz, J.H. (1993) *supra*; Sacktor et al. (1993) *supra*.

- PKM is usually thought to be produced by  
25 limited proteolysis of PKC, separating the enzyme's regulatory and catalytic domains. This may occur early after a high-frequency tetanus. Recent evidence shows, however, that the long-lasting PKM $\zeta$  may also be derived from a brain-specific mRNA that encodes only the  
30 catalytic domain of  $\zeta$ . Andrea et al. (1995) Biochem. J.

310:835-843; Powell et al. (1994) Cell Growth Differ.  
5:143-149.

PKC is a family of multifunctional protein  
kinases, first discovered by Nishizuka in 1977. Takai et  
5 al. (1977) J. Biol. Chem. 252:7603-7609; Inoue et al.  
(1977) J. Biol. Chem. 252:7610-7616. PKC consists of two  
domains separated by a hinge region: an amino-terminal  
regulatory domain, which contains an autoinhibitory  
pseudosubstrate sequence and second messenger/lipid  
10 binding sites, and a carboxy-terminal catalytic kinase  
domain. PKC is held in an inactive state in the cytosol  
by the interaction between the regulatory and catalytic  
domains. When there is an increase in lipid second  
messengers (or, for some isoforms,  $\text{Ca}^{2+}$ ), PKC translocates  
15 from the cytosolic to membranous (or cytoskeletal)  
compartments, and a change in its conformation occurs,  
displacing the regulatory from the catalytic domain,  
releasing the autoinhibition, and activating the enzyme.  
The 10 different forms of PKC are divided into 3 groups:  
20 conventional ( $\alpha$ ,  $\beta\text{I}$ ,  $\beta\text{II}$ ,  $\gamma$ ), novel (or new,  $\delta$ ,  $\epsilon$ ,  $\eta$ ,  $\theta$ ),  
and atypical ( $\zeta$ ,  $\iota/\lambda$ ), each of which is activated by a  
distinct set of second messengers. (PKD or PKC $\mu$  is a  
PKC-related molecule with a catalytic domain closer to  
CaM-kinase). The conventional PKCs are activated by  $\text{Ca}^{2+}$   
25 and diacylglycerol (DAG); the novel by DAG, but not  $\text{Ca}^{2+}$ ;  
and the atypical by neither DAG or  $\text{Ca}^{2+}$ , but by alternate  
lipid-second messengers, including arachidonic acid,  
phosphoinositide 3-OH kinase products, and ceramide.

A second mechanism for permanently activating  
30 PKC, also discovered by Nishizuka, is the cleavage by

calpain or their proteases at the hinge region, to permanently separate the regulatory from the catalytic domains. The independently active kinase domain is called PKM. ("M" stands for  $Mg^{2+}$ , although this requirement turned out to be for the  $Mg^{2+}$  in  $Mg^{2+}$ -ATP). PKM formation results in a persistently active kinase and is not the typical way PKC is activated. It has been found that stable PKM formation occurs endogenously only for a single isoform,  $\zeta$ , and only in brain. Naik et al. (submitted for publication). Recently, PKM $\zeta$  has also been reported in a neuronally differentiated cell line. Oehrlein et al. (1998) Eur. J. Cell. Bio. 77:323-337.- Stable PKM forms for the other isoforms have been observed only in pathological conditions: PKM $\epsilon$  in breast cancer tumor cells (Baxter et al. (1992) J. Biol. Chem. 267: 1910-1917) and heart ischemia (Urthaler et al. (1997) Cardiovasc. Res. 35:60-67) and PKM $\delta$  in apoptosis (Emoto et al. (1996) Blood 97:1990-1996; Denning et al. (1998) J. Biol Chem. 273:29995-30002).

Protein kinase M zeta (PKM $\zeta$ ) is a form of protein kinase C which has a fundamental role in the formation and maintenance of memory. PKM $\zeta$  is a critical molecule in the most widely-studied physiological model of memory, long-term potentiation (LTP) of synapses (Sacktor, et al., (1993) *supra.*; Osten, et al., (1996) J. Neurosci. 16(8):2444-2451; Hrabetova and Sacktor, (1996) J. Neurosci. 16(17):4324-5333).

#### **SUMMARY OF THE INVENTION**

The present invention provides methods and compositions for enhancing memory and treating brain and

spinal cord injury by the administration of a therapeutically effective amount of one or more atypical forms of protein kinase C (PKC) such as PKM $\zeta$  or PKC iota/lambda.

5           In one aspect, the present invention provides method of enhancing synaptic transmission in an animal comprising the administration of a therapeutically effective amount of one or more atypical forms of PKC such as PKM $\zeta$  or PKC iota/lambda.

10           In another aspect, the present invention provides a method of maintaining memory in an animal comprising the administration of a therapeutically effective amount of one or more atypical forms of PKC such as PKM $\zeta$  or PKC iota/lambda.

15           In still another aspect, the present invention provides a method for enhancing synaptic transmission or maintaining memory comprising the administration of DNA encoding the human (or animal) sequence of PKM $\zeta$ . In yet another aspect, the present invention provides a method  
20 for causing amnesia or decreasing synaptic transmission, comprising the administration of a therapeutically effective amount of a PKM $\zeta$  inhibitor. Uses for decreasing synaptic transmission include, for example, the treatment of acute or chronic pain, treatment of drug  
25 or alcohol addiction, and treatment of epilepsy.

          In still yet another aspect, the present invention provides a method for causing amnesia or decreasing synaptic transmission, comprising the administration of a therapeutically effect amount of a  
30 dominant negative PKM $\zeta$  inhibitor, DNA encoding the human (or animal) sequence of dominant negative PKM $\zeta$ , or

antisense to PKM $\zeta$ .

In another aspect, the present invention provides a method of causing amnesia in an animal comprising the administration of a therapeutically effective amount of a PKM $\zeta$  inhibitor or a PKC iota/lambda inhibitor.

In still another aspect, the present invention provides a pharmaceutical composition comprising PKM $\zeta$  and a pharmaceutically acceptable carrier.

In yet another aspect, the present invention provides a pharmaceutical composition comprising PKC iota/lambda and a pharmaceutically acceptable carrier.

#### **BRIEF DESCRIPTION OF THE DRAWINGS**

Figure 1A shows individual EPSC responses to injected PKM $\zeta$ .

Figure 1B shows the time course of EPSC amplitudes.

Figure 2A shows whole cell recording of CA1 pyramidal cells with 3nM PKM $\zeta$ . Increases in synaptically evoked AMPA/kainate responses with the introduction of PKM $\zeta$  were reversible by chelerythrine (1 $\mu$ M).

Figure 2B shows the time course of EPSC amplitudes.

Figure 3 shows a silver stain of purified recombinantly expressed PKM $\zeta$ .

Figure 4 shows specificity of inhibition of PKM $\zeta$  relative to other protein kinases by chelerythrine. PKCA is PKC alpha phosphorylation of myelin basic protein (MBP). PKCE is PKC epsilon phosphorylation of MBP. PKM is PKM $\zeta$  phosphorylation of MBP. CKII is Ca<sup>2+</sup>-calmodulin-dependent protein kinase II phosphorylation of Syntide®

(Calbiochem, San Diego, CA.) peptide substrate.

Figure 5 shows that a myristolated zeta inhibitory pseudosubstrate peptide (1 $\mu$ M), reverses LTP maintenance.

5           Figure 6 shows the DNA sequence (derived from cDNA) encoding human PKM $\zeta$ .

Figure 7 shows postsynaptic exposure to dominant negative inhibitor PKM $\zeta$ -K281W (20 nM) prevents LTP. (A) Silver stain of baculovirus/Sf9-expressed PKM $\zeta$ -K281W protein placed into whole-cell recording pipette. PKM $\zeta$ -K281W appears as a doublet. (B) Depolarization to -40 mV at 1 and 10 min after diffusion of PKM $\zeta$ -K281W shows no obvious effect on synaptic responses mediated by AMPA and NMDA receptors. (C) Upper traces, whole-cell EPSCs pre-and posttetanization, showing no persistent potentiation after exposure to PKM $\zeta$ -K281W. Lower traces, simultaneously recorded field potentials show LTP. (D) Time course of whole-cell recording with PKM $\zeta$ -K281W. Tetanization shows only PTP. Inset, time course of simultaneous field recordings shows LTP.

#### **DETAILED DESCRIPTION OF THE INVENTION**

The present invention provides methods and compositions for enhancing memory and treating brain and spinal cord injury by the administration of a therapeutically effective amount of one or more atypical forms of PKC. In a preferred embodiment an atypical form of PKC is PKM $\zeta$ . In another preferred embodiment an atypical form of PKC is PKC iota/lambda. In accordance with the present invention it has been determined that PKM $\zeta$  is both necessary and sufficient for the long-term



1  
maintenance of LTP. Moreover, it has been determined in  
accordance with the present teachings that the function  
of PKM $\zeta$  is to store and consolidate memories in the  
brain.

5           In accordance with the present invention,  
members of the class of compounds known as atypical forms  
of PKC such as protein kinase M zeta (PKM $\zeta$ ) and PKC  
iota/lambda have been found to maintain or consolidate  
10 long term changes in synaptic strength in vertebrates,  
the mechanism for long term memory. The present  
invention elucidates PKM $\zeta$ 's role in maintaining enhanced  
synaptic transmission with studies of long-term  
potentiation (LTP). Conversely, inhibition of PKM $\zeta$  may  
15 cause amnesia, which may be useful in the treatment of  
traumatic stress disorders, phobias and acute or chronic  
pain.

Other agents that have been proposed to enhance  
memory are essentially stimulates (like coffee) or agents  
designed to enhance the induction of long-term  
20 potentiation (LTP)-like processes (such as drugs to  
increase cAMP). PKM $\zeta$  is the first molecule whose  
function is to maintain memories in vertebrates. In  
accordance with the present invention, when PKM $\zeta$  is  
injected into neurons it persistently enhances synaptic  
25 transmission.

In one embodiment the present invention  
contemplates a method of treating a brain injury in an  
animal comprising the administration of a therapeutically  
effective amount of one or more atypical forms of PKC  
30 such as PKM $\zeta$  or DNA encoding PKM $\zeta$  message. By  
therapeutically effective amount is meant an amount of an

atypical form of PKC high enough to significantly positively modify the condition to be treated but low enough to avoid serious side effects (at a reasonable benefit/risk ratio), within the scope of sound medical judgment. For example, a therapeutically effective amount of PKM $\zeta$  will vary with the particular injury being treated, the age and physical condition of the patient being treated, the severity of the injury, the duration of treatment, the nature of concurrent therapy and the specific PKM $\zeta$  employed.

In another embodiment the present invention contemplates a method of treating a spinal cord injury in an animal comprising the administration of a therapeutically effective amount of one or more atypical forms of PKC such as, for example, PKM $\zeta$ .

In still another embodiment, the present invention contemplates a method of enhancing synaptic transmission in an animal comprising the administration of a therapeutically effective amount of one or more atypical forms of PKC such as, for example, PKM $\zeta$ .

The present invention also contemplates a method of inducing amnesia in an animal by the administration of a therapeutically effective amount of a PKM $\zeta$  inhibitor. In preferred embodiments the PKM $\zeta$  inhibitor is chelerythrine, myristolated zeta inhibitory pseudosubstrate (MZIP) peptide (myr-Ser-Ile-Tyr-Arg-Arg-Gly-Ala-Arg-Arg-Trp-Arg-Lys-Leu-OH), or dominant negative form of PKM $\zeta$  such as, for example, PKM $\zeta$ -K281W, or antisense to PKM $\zeta$  mRNA. MZIP has an IC<sub>50</sub> of 10-100nM for PKM $\zeta$  and 10,000nM for PKC gamma and therefore is a more specific inhibitor than chelerythrine (see Fig. 5).

Candidates for the induction of selective amnesia contemplated by the present invention are preferably humans having, for example, post-traumatic stress disorders and phobias.

5           The present invention also contemplates a method of reducing synaptic transmission in selective areas of the brain or spinal cord by the administration of a therapeutically effective amount of PKM $\zeta$  inhibitor. Candidates for the reduction of synaptic transmission  
10 contemplated by the invention are preferably humans having, for example, disorders of pain, drug or alcohol addiction, or excess neuronal activity as in epilepsy.

Still another embodiment of the present invention contemplates pharmaceutical compositions  
15 containing one or more atypical forms of PKC such as, for example, PKM $\zeta$ .

The active ingredients of a pharmaceutical composition containing PKM $\zeta$  or a nucleic acid encoding PKM $\zeta$  is contemplated to exhibit effective therapeutic  
20 activity, for example, in enhancing memory, and treating brain and spinal cord injuries. Thus the active ingredients of the therapeutic compositions containing PKM $\zeta$  is administered in therapeutic amounts which depend on the particular disease. For example, final  
25 concentrations of PKM $\zeta$  in brain or spinal cord to be achieved by administration may be about 1 nanomolar. The dosage regimen may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily or the dose may be  
30 proportionally reduced as indicated by the exigencies of the therapeutic situation. Administration of one or more

atypical forms of PKC such as, for example, PKM $\zeta$  into the brain or spinal cord may be intracranially or intrathecally, i.e., by intrathecal pump or repository. Depending on the route of administration, the active ingredients which comprise PKM $\zeta$  may be required to be coated in a material to protect said ingredients from the action of acids and other natural conditions which may inactivate said ingredients.

For example, PKM $\zeta$  may be administered in an adjuvant or in liposomes. Adjuvants contemplated herein include resorcinols, non-ionic surfactants such as polyoxyethylene oleyl ether and n-hexadecyl polyethylene ether. Liposomes include water-in-oil-in-water P40 emulsions as well as conventional liposomes.

Under ordinary conditions of storage and use, the preparations of the present invention contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, liquid polyethylene glycol, and the like), suitable mixtures thereof and vegetable oils. The proper fluidity can be maintained,

for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of

5 microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases it will be preferable to include isotonic agents, for example,

10 sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

15 Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are

20 prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable

25 solutions, the preferred methods of preparation are vacuum-drying and the freeze-drying technique which yield a powder of the active ingredient plus any additional desired ingredient from previously sterile-filtered solution thereof.

30 It is especially advantageous to formulate compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form

as used herein refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of the active material calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the novel dosage unit forms of the invention are dictated by and directly depending on (a) the unique characteristics of the active material and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such as active material for the treatment of injury in living subjects having a condition in which bodily health is impaired as herein disclosed in detail.

The principal active ingredient is compounded for convenient and effective administration in effective amounts with a suitable pharmaceutically acceptable carrier in dosage unit form as hereinbefore disclosed. A unit dosage form can, for example, result in achieving, for example, about 0.1 to about 10 nanomolar concentrations of PKM $\zeta$  in the brain or spinal cord.

As used herein "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and adsorption delaying agents, and the like. The use of such media agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, use thereof in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

Administration of an atypical form of PKC, such as PKM $\zeta$  may also include altered forms or derivatives of PKM $\zeta$  or drugs that enhance its activity, stability, or accessibility to the nervous system. The identification of applicable PKM $\zeta$  enhancing drugs are readily tested or screened by examining the effects of drugs or PKM $\zeta$ 's phosphorylation in vitro (as measured in Figure 4) or on PKM $\zeta$ 's effect on synaptic transmission when injected into neurons as in Figure 2.

Administration of PKM $\zeta$  DNA into brain or spinal cord may also be by gene-transfer technology. Such technologies include, but are not limited to, viruses, liposomes, and altered forms or derivatives of DNA or RNA.

Administration of inhibitors of PKM $\zeta$  activity include drugs, such as chelerythrine, myristolated zeta inhibitory pseudosubstrate peptide and altered forms of PKM $\zeta$  that, through dominant negative effects inhibit endogenous PKM $\zeta$ 's activity or effectiveness. Such dominant negative agents include, but are not limited to, inactive forms or portions of PKM $\zeta$ . Inhibition of PKM $\zeta$  function may also include decreasing levels of endogenous PKM $\zeta$  through administration of antisense to the sequence of PKM $\zeta$ .

The following Examples serve to further illustrate the invention without in any way limiting same.

### EXAMPLE 1

The effect of the increase of PKM $\zeta$  on synaptic transmission.

#### 5 Intracellular perfusion of PKM $\zeta$ .

PKM $\zeta$ , purified from a baculovirus/Sf9 expression system to near homogeneity, was perfused into a CA1 pyramidal cell through a whole cell recording pipette (Fig. 1A-1B). The concentration of PKM $\zeta$  was  
10 about 0.8 nM PKM $\zeta$ , with 0.03 pmol/min/ $\mu$ l activity. In order to isolate AMPA/kainate responses, the cell, recorded at the soma, was voltage-clamped at the IPSC reversal potential ( $\sim$ -70) and Cs-gluconate-based electrode solution was used (to block GABA $_B$  effects).  
15 Stimulation was every 10 sec in the radiatum, and input resistance was monitored and did not change throughout the experiment. Over 5-10 min, there was  $\sim$ 60% increase in AMPA/kainate EPSCs, which then stabilized. In contrast, inactivated PKM $\zeta$  had no effect. In comparison  
20 with the concentration of autoactive thiophosphorylated CaM-kinase II reported to potentiate synaptic responses in CA1 pyramidal cells by similar whole cell techniques, the findings on the potentiation of AMPA/kainate currents indicated that PKM $\zeta$  was 200- to 1000-fold more potent  
25 than CaM-kinase II.

#### Electrophysiological recording.

Hippocampal slices were prepared from Sprague-Dawley rats as described in Example 2. Patch electrodes  
30 were pulled (two-stage) from 15 mm O.D. borosilicate glass (World Precision Instruments, Sarasota, FL) on a Narishige PP-83 vertical puller. Recording pipettes had



tip resistances of 2-5 M $\Omega$  and contain (in mM): Cs-gluconate or K-gluconate 130-; MgCl<sub>2</sub>, 2; CaCl<sub>2</sub>, 2; EGTA, 10; HEPES, 10; Na-ATP, 2; pH adjusted to 7.25 with either CsOH or KOH. This mixture had been shown to reduce the rundown of GABA<sub>A</sub>-receptor mediated responses. Cesium was used to block potassium currents, including slow GABA<sub>B</sub> IPSCs. Electrode solutions included QX-314 (10mM) to block Na<sup>2+</sup> currents to prevent cell spike discharges at depolarized holding potentials. Whole-cell recordings were obtained from CA1 pyramidal cells using blind patch techniques. The recording pipette was slowly advanced through the tissue, with brief voltage steps (-10mV, 10 ms) applied to monitor electrode resistance. Once a deflection in the electrode's current response was detected signifying contact with the membrane of the target cell soma, slight negative pressure was applied to form a cell-electrode seal of >1 G $\Omega$ .

Membrane breakthrough was accomplished with either additional suction or current pulses. Following membrane rupture, 2 min of settling time was allowed before formal recording. Voltage steps (-10 mV) were applied to monitor the access resistance and input capacitance. Signals were recorded under voltage-clamp with an Axoclamp 2A amplifier (Axon Instruments, Foster City, CA). Resting input resistance was measured from the current response to a -10 mV voltage step from holding potential (usually resting membrane potential, ~-60 mV with the intracellular solution). Cells were accepted for study only if resting input resistances of >100 M $\Omega$  and access resistances <20 M $\Omega$  were observed.

If cell access resistance increased significantly during the course of the recording (>20%),

the data was discarded. Data signals were digitized at 94 kHz via a 14-bit PCM interface (VR-10B Digital Data Recorder, Instrutech Corp., Element, NY) and stored on VHS tape for later analysis with pCLAMP software (Axon Instruments) on an IBM-compatible Pentium-II microcomputer.

Synaptic events were evoked by extracellular stimulation with bipolar, coated tungsten electrodes placed in stratum radiatum lateral to the recording electrode. Cathodal shocks (2-10 V; 200  $\mu$ s duration) were delivered through a digitally controlled stimulus isolation unit (World Precision Instruments) at a low frequency (0.1 Hz). Unless otherwise noted, drugs were delivered in the bath.

15

## EXAMPLE 2

Methods: Preparation of hippocampal slices. After anesthesia with halothane, transverse 450 $\mu$ m slices were prepared from 3-4 week old Sprague-Dawley rats with a  
5 McIlwain tissue chopper. During the dissection, the hippocampus was kept cold with multiple washes of a dissection saline at 4°C (125 mM NaCl, 2.5 mM KCl, 1.25 mM NaH<sub>2</sub>PO<sub>4</sub>, 26 mM NaHCO<sub>3</sub>, 11 mM glucose, 10 mM MgCl<sub>2</sub> and 0.5 mM CaCl<sub>2</sub>, pH 7.4, equilibrated with 95% O<sub>2</sub>/5% CO<sub>2</sub>).

10 The slices were transferred to an interface chamber (Fine Science Tools) for incubation in the dissection saline for 1 hour at 32°C. The divalent ion concentrations were then changed to 1.2 mM MgCl<sub>2</sub> and 1.7 mM CaCl<sub>2</sub> (physiological saline).

15

Intracellular perfusion of PKM $\zeta$ .

Analysis of the effect of PKM $\zeta$  on evoked EPSCs in CA1 pyramidal neurons and effects of chelerythrine.

The effects of PKM $\zeta$  cell injection on  
20 excitatory synaptic currents were assessed by including the kinase in the whole-cell pipette and measuring EPSC amplitude over time. Synaptic events were recorded in the soma of hippocampal CA1 pyramidal cells with Cs-gluconate-based electrode solutions containing varying  
25 concentrations of PKM $\zeta$  (0.1, 1, 10 nM). With cells voltage-clamped at the IPSC reversal potential ( $E_{\text{ipsc}}$ ,  $\sim$ -70 mV), isolated ensemble EPSCs were elicited by low-frequency (0.1 Hz) electrical stimulation applied in stratum radiatum. Stimulus intensity was set at moderate  
30 levels ( $\sim$ 5 V) to selectively evoke fast EPSCs (i.e., AMPA/kainate-mediated) and was held constant throughout the experiment. Control data consisted of measurements

of EPSC amplitude and duration immediately following establishment of a whole-cell recording (i.e., before PKM $\zeta$  diffusion into the target cell), and parallel experiments with the kinase denatured by boiling or  
5 inactivated by multiple freeze/thaw. Activity of the perfused PKM $\zeta$  was determined by phosphorylation assay *in vitro* with each experiment.

EPSCs was continuously monitored to assess the effects of intracellular PKM $\zeta$  on peak EPSC magnitude and  
10 duration. Periodically, current-voltage records were taken to check constancy of the EPSC reversal potential ( $E_{EPSC}$ ), and the input resistance of the cell was monitored throughout the experiment. Application of the non-NMDA-receptor antagonist CNQX (10 $\mu$ M) was applied to slices to  
15 confirm that EPSCs elicited at  $E_{IPSC}$  with moderate stimulus intensity were mediated solely through non-NMDA receptors.

Once an effect was observed and appeared stable, the competitive PKC catalytic domain inhibitor, chelerythrine, was added to the bath to attempt to  
20 reverse the effect by preventing PKM $\zeta$ 's phosphorylation. The drug was then washed out. An example of an experiment with chelerythrine using 3 nM PKM $\zeta$  is provided in Figures 2A-2B.

25

### EXAMPLE 3

#### Purification of baculovirus-expressed PKM $\zeta$ from Sf9

cells: Spodoptera frugiperda (Sf9) cells were grown in SF-900 II SFM insect cell culture medium (Gibco)

- 5 containing 5 $\mu$ g/ml gentamicin. To express the PKM $\zeta$ ,  
10x10<sup>8</sup> cells of a healthy, log phase Sf-9 culture were  
spun, resuspended in 125 ml of medium, and infected with  
25 ml of the baculovirus- $\zeta$  virus stock (gift of Sylvia  
Stable). Following a 0.5 hr incubation at room  
10 temperature, additional medium was added to the cells,  
which were then seeded at a density of 1x10<sup>6</sup> cells/ml.  
After 3 days, the cells were spun, washed with PBS, and  
then homogenized in 65 ml of homogenization buffer. A 2-  
step purification, employing DEAE Fast Flow Sepharose and  
15 Superdex 75 (preparation grade, Pharmacia) columns, was  
performed to purify baculovirus-expressed PKM $\zeta$ . Activity  
of the PKM $\zeta$  was assayed on the same day as each whole  
cell experiment. Silver stain of baculovirus,  
recombinantly expressed PKM $\zeta$  showing purity of  
20 preparation. (See Fig. 3)

#### EXAMPLE 4

##### Specificity of chelerythrine as inhibitor of PKM $\zeta$

- 5 Inhibition by the drug chelerythrine of phosphorylation of exogenous substrates by various protein kinases showed that chelerythrine was greater than 10-fold selective as an inhibitor of PKM $\zeta$  activity. (See Figure 4).

### EXAMPLE 5

#### Sense, antisense, and amino-acid sequence of human PKM $\zeta$

Human sequence, obtained from published  
expressed sequence tags, were obtained by analogy with  
5 rat sequence (Ono, et al., 1988), which was identified as  
containing the open reading frame of PKM $\zeta$ . The PKM $\zeta$   
sequence is used for gene transfer technology to increase  
levels of PKM $\zeta$  in the nervous system. Dominant negative  
inhibitors of PKM $\zeta$  are obtained by changing or deleting  
10 sequences in the ATP-binding site or by administering  
selective domains of the protein, e.g., the carboxy-  
terminal domain.

Dominant negative PKM $\zeta$  was obtained by  
elimination or alteration of ATP-binding domain (marked),  
15 and by administration of carboxy-terminal domain  
(marked). Antisense was achieved by administration of  
all or part of antisense sequence to PKM $\zeta$ . (See Figure  
6).

### EXAMPLE 6

In order to confirm that PKM $\zeta$  mediates potentiation of synaptic transmission during LTP, a dominant negative inhibitory form of PKM $\zeta$  (PKM $\zeta$ -K281W) was introduced postsynaptically into a CA1 pyramidal cell its effect on LTP was examined. The lysine<sup>281</sup> to tryptophan mutation in the catalytic domain of PKM $\zeta$ -K281W) abolished kinase activity by disrupting ATP-binding. Twenty nanomolar of the dominant negative inhibitor (Fig. 7A) was included in a whole-cell recording pipette and allowed to diffuse for 10 min into a CA1 pyramidal cell, voltage clamped at -75 mV to isolated synaptic AMPA responses. PKM $\zeta$ -K281W had no obvious effect on synaptic transmission mediated by AMPA and NMDA receptors, sampled at -40 mV at 1 min and 10 min (Fig. 7B). PKM $\zeta$ -K281W completely eliminated persistent synaptic potentiation in the cell, but not post-tetanic potentiation (PTP) (Fig. 7C, upper traces, D, large graph). Twenty minutes after the tetanic stimulation, the EPSCs were 101.6 $\pm$ 1.9% of baseline responses (n=4). Simultaneous LTP in the field responses of the slice was observed (Fig. 7C, lower traces, and D, inset graph).

PKM $\zeta$ -K281W was expressed using the MaxBac 2.0 Baculovirus/Sf9 system (Invitrogen, Carlsbad, CA). An insert containing amino acids 158-592 of PKC $\zeta$  (Drier, et al. (2000) 30<sup>th</sup> Annual Meeting of the Society of Neuroscience, New Orleans, LA, generous gift from Jerry Yin, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY) was subcloned into the baculovirus transfer vector pBlueBacHis2B between the EcoRI and SalI sites. The construct was sequenced for verification, and then cotransfected into Sf9 cells with linearized baculovirus



genome. Recombinant protein was expressed in Sf9 cells, and purified using a Ni<sup>2+</sup> column (Invitrogen, Xpress<sup>TM</sup> Purification Kit, Carlsbad, CA), and analyzed by silver stain (Fig. 7A) and Western blot (data not shown).